



Warner Instruments
Epithelial Voltage Clamp
Models EC-800 & EC-800LV



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The **EC-800** and **EC-800LV** Epithelial Voltage Clamps from Warner Instruments provide accurate measurements of transepithelial voltage, short circuit current and membrane resistance. Important features include fluid resistance compensation, membrane resistance readout, high voltage compliance and small, water tight headstages. Operating modes include voltage clamp, current clamp, voltmeter and resistance.

Unique Features Include

- ✓ **Membrane Resistance Measurement:** Accurate resistance measurements are made with the membrane mounted in the chamber. The measurement is made using a low frequency bipolar signal to avoid polarization of the membrane [ideal for monolayers]. Resistances up to 200 k Ω are displayed on the meter with push button convenience.
- ✓ **Clamp Speed Selection up to 10 μ s:** Three clamp speeds provide optimum recording conditions for a variety of applications. In Fast mode, preparations with low access resistance (small tissues or monolayers) can be clamped with speeds up to 10 μ s. Typical Ussing chambers with larger tissues will use Medium or Slow modes for stable, oscillation free clamping.
- ✓ **Independent Voltage and Current Commands:** Internal command controls are provided for both voltage clamp and current clamp modes.
- ✓ **Watertight Headstage with Model Membrane:** The small, compact headstage can be located close to the measurement site to keep input leads short for reduced noise pick-up. The model membrane circuit simulates a preparation to provide convenient operational checks of the clamp. Internal circuits are protected against the invasion of corrosive saline solutions by a watertight seal.
- ✓ **Choice of Compliance Voltage:** The high voltage compliance of the **EC-800** (± 120 V) is important for studies of low resistance [leaky] epithelial cells and in applications in which long agar leads in the current passing circuit produce large voltage drops which must be compensated. Additionally, high compliance aids in charging large membrane capacitances typical of epithelial tissues, resulting in faster settling times and improved overall clamp performance. Studies with small tissue samples or monolayers in set-ups with low access resistance may not require high compliance. For these applications, the **EC-800LV** (± 15 V) offer both a lower cost and a safer environment for the membrane.

Additional Features Include

- ✓ **High CMR:** Differential voltage recordings are made with very high common mode rejection providing accurate measurements free from the effects of common mode potential changes of a noisy environment.
- ✓ **External Instrument Control:** The clamp can be operated by an external programmer, lab timer or computer. Logic control of clamp mode and clamp command levels is possible as well as simultaneous mixing of external linear commands.

NOMENCLATURE

Text conventions

This manual refers to amplifier controls at three functional levels; control blocks, specific controls within a block, and settings of specific controls. To minimize the potential for confusion, we have employed several text conventions which are specified below. Since our goal is to provide clarity rather than complexity, we welcome any feedback you may wish to provide.

- Warner Instrument product numbers are presented using **bold type**.
- References to instrument panel control blocks are specified using UNDERLINED SMALL CAPS. (e.g. METER, CLAMP COMMANDS)
- References to specific controls within a block are specified using NON-UNDERLINED SMALL CAPS. (e.g. MODE SWITCH, TIMER RANGE)
- References to individual control settings are specified in *italic type*. (e.g. *Amplify, 100 mV*)
- Input connections to the headstage are printed in a **bold type**. (e.g., **I1 output, V2 input**)
- Special comments and warnings are presented in **highlighted text**.

Any other formatting should be apparent from context.

**THIS EQUIPMENT IS NOT DESIGNED NOR INTENDED
FOR USE ON HUMAN SUBJECTS**

CONTROL DESCRIPTION

The following is a description of the operating controls, inputs and outputs located on the front and rear panels of the instrument.

Front panel control blocks

The front panel is comprised of four labeled control blocks termed INPUT OFFSET, FLUID RESISTANCE, MODE, and COMMANDS. A fifth, unmarked section is termed the METER control block.

Input Offset

The toggle switch selects input polarity (+ or -) or *off* (no offset). The ten turn control provides input offset adjustment from 0-120 mV. The HEADSTAGE connects to the amplifier in this control block.

Fluid Resistance and monitors



This control block is used to compensate for any voltage drops due to the intrinsic resistance of the bathing solutions. A 25 μA current (this value can be adjusted internally) is injected at the HEADSTAGE I1 output. The measurement/adjustment is made by depressing the PUSH TO ADJUST button and turning the ten turn dial until *zero* is read on the digital meter. The fluid resistance is read from the dial (0-100 Ω).

NOTE: This range can be extended to 1 kΩ, see the Appendix for details.

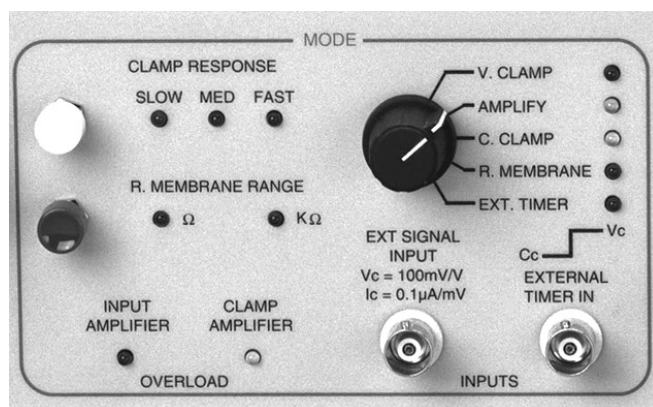
Monitors - Two output BNC's are included in this control block for communicating amplifier outputs to data visualization and storage devices and are described below.

<i>V Monitor x10</i>	Output is the differential voltage (V1-V2) multiplied by 10.
<i>I Monitor 10mV/μA</i>	Output voltage is proportional to the current flowing through the preparation.

Mode

This control block represents the main control section on the front panel and is comprised of a five position MODE SELECTOR switch, various LED indicator lights, the CLAMP RESPONSE push button, the R MEMBRANE RANGE push button, and two INPUTS BNC's.

The MODE SELECTOR SWITCH is used to select among the various operational modes available in the instrument.



Available selections are *V Clamp*, *Amplify*, *C Clamp*, *R Membrane*, and *Ext Timer*. LED's indicate the MODE selection. The meaning of MODE selections are shown in the table below.

<i>V Clamp</i>	Selects voltage clamp mode.
<i>Amplify</i>	With the exception of <i>fluid resistance</i> measurements, the clamp amplifier is disconnected from the headstage resulting in no signal at the <u>HEADSTAGE I1 output</u> . This is a true amplify operation.
<i>C Clamp</i>	Selects current clamp mode.
<i>R Membrane</i>	Selects membrane resistance measurement mode. A constant current, 2 Hz square wave is injected and the resultant voltage is measured across the membrane. The calculated resistance is displayed on the meter.
<i>Ext Timer</i>	External timer mode. A TTL signal applied to the EXTERNAL TIMER IN BNC will switch the amplifier between current clamp (TTL= <i>high</i>) and voltage clamp (TTL= <i>low</i>) modes.

The CLAMP RESPONSE push button selects *fast*, *med* or *slow* clamp speeds. This control is active in both current and voltage clamp modes.

The R MEMBRANE RANGE push button selects the resistance measurement range (Ω or $k\Omega$) used when *R Membrane* is selected on the MODE SELECTOR switch. A constant current bipolar square wave is passed through the membrane (10 μ A in the 0-2 $k\Omega$ range and 1 μ A in the 0-100 $k\Omega$ range). The membrane resistance read on the panel meter which scales automatically with the range selected.

Two input BNC's are included within this control block and are described below.

EXT SIGNAL INPUT: An external voltage or current clamp command input. Applied current or voltage commands are summed with their respective counterparts in the COMMANDS control block. In voltage clamp mode, the ratio of EXT SIGNAL INPUT voltage to applied clamp voltage is 0.01 V/mV. In current clamp mode the ratio of EXT SIGNAL INPUT voltage to applied clamp current is 10 mV/ μ A. This input is inactive when GATE INPUT is *high*.

EXTERNAL TIMER IN: Activated by selecting *Ext Timer* in the MODE control block. A TTL signal at this BNC switches the amplifier between voltage clamp (TTL=*low*) and current clamp (TTL=*high*) modes.

Finally, two LED's are provided to indicate overload conditions in the clamp amplifier or input amplifier circuitry.

Commands

The VOLTAGE control, comprised of a 10-turn potentiometer and a three position toggle switch, adjusts the holding potential effective in voltage clamp mode. The operational range is adjustable from zero to ± 100 mV with the polarity selected by the toggle switch.

The current control, is also comprised of a 10-turn potentiometer and a three position toggle switch. This control adjusts the holding current effective in current clamp mode and the operational range is adjustable from zero to ± 1 mA with the polarity selected by toggle switch.

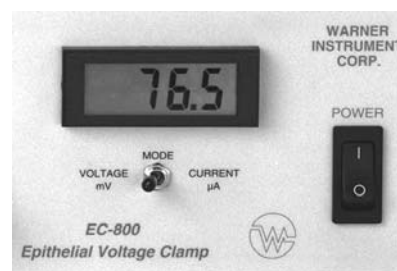
NOTE: A TTL=*high* logic level at the GATE INPUT BNC (located on the instrument rear panel) will disable the COMMANDS control block.



Meter

The METER section is comprised of a 3.5 digit LCD, a 2 position toggle switch, and the power switch for the unit.

The METER toggle switch (MODE) selects between *current* (μA) and *voltage* (mV) readings. Full scale is $\pm 1,900$ μA and ± 199 mV for *current* and *voltage* positions, respectively. Either current or voltage can be displayed for any selection within the MODE control block, with the exception of R MEMBRANE mode.



In R MEMBRANE mode, the meter reads the membrane resistance in either Ω or k Ω , as selected by the R MEMBRANE RANGE push button in the MODE control block.

Rear panel description

The instrument rear panel provides several connections for control input and data output.

Line power connector and fuse

The power cord connects to the power connector. Check the serial number tag on the rear panel to be sure that the instrument is wired for the operating voltage used. The fuse (3AG type, regular blow) used in the **EC-800** will depend on the line voltage; 1/2 amp for 100 or 130 VAC, and 1/4 amp for 220 or 240 VAC. Replacement should only be made with the proper ampere rating.

I/O port

This 9 pin connector allows the **EC-800** to be connected for computer operation. Pin designations are:

1	V MONITOR x 10	5	EXTERNAL TIMER IN
2	I MONITOR	6, 7 & 8	CIRCUIT GROUND
3	EXT SIGNAL INPUT	9	no connection
4	GATE INPUT		

Gate input

Used to control the behavior of the EXTERNAL TIMER IN input BNC in the MODE control block. A TTL=*low* signal at the GATE INPUT BNC instructs the EXTERNAL TIMER IN BNC to select between the values set for voltage or current clamp modes. By comparison, a TTL=*high* signal at the GATE INPUT BNC instructs the EXTERNAL TIMER IN BNC to select for *zero current clamp* or for *zero voltage clamp* for current clamp mode or voltage clamp mode, respectively.

All clamp commands (current, voltage and external) are deactivated with a TTL=*high* signal applied to this BNC.

Grounds

Both circuit (black) and chassis (green) grounds are provided at rear binding posts. The instrument is shipped with the two grounds connected *via* a shorting bar. If needed, this shorting bar can be disconnected allowing separation of the grounds. Separating these grounds can, in some cases, result in a lowering of the 60 Hz noise level, however, experimentation will determine which is best for each set up.

Additional components and comments

Connecting to line power

The model EC-800 is supplied with a 3-conductor line cord. One conductor provides a connection between the instrument housing and the power system earth ground. Safe operation of this instrument will be assured provided that the power outlet is wired correctly and is connected to earth. If the ground pin of the line cord is removed for any reason the instrument chassis must be connected to earth ground using a separate heavy gauge (14 or larger) ground wire.

High voltage outputs

CAUTION: The clamp is capable of high power output (± 120 V @ 10 mA). When handling a HEADSTAGE I1 electrode cable, be sure to set the MODE SELECTOR switch to *amplify* and the HEADSTAGE SELECTOR switch to *off*.

Headstage

The headstage has inputs for the **I1**, **V1**, **V2**, and **I2** **electrode cables** and a three position switch selecting *on*, *off* or *test* modes. In addition, *test* mode has a two position toggle switch selecting for *Vmem* or *Fluid Resistance*.

Moreover, the headstage is equipped with a built-in model membrane allowing for a rapid and convenient instrument test. (See page 14 for test procedure).



Connecting the headstage

The headstage connects to the instrument via an 8-pin DIN connector in the INPUT OFFSET control block. To reduce noise the headstage should be placed as close to the test site as possible.

SETUP AND INITIAL TEST

The purpose of this section is twofold. First, a description of the equipment needed to measure the basic electrical properties of an epithelium is provided. Second, a series of procedures are provided to test the performance of the instrument.

Equipment

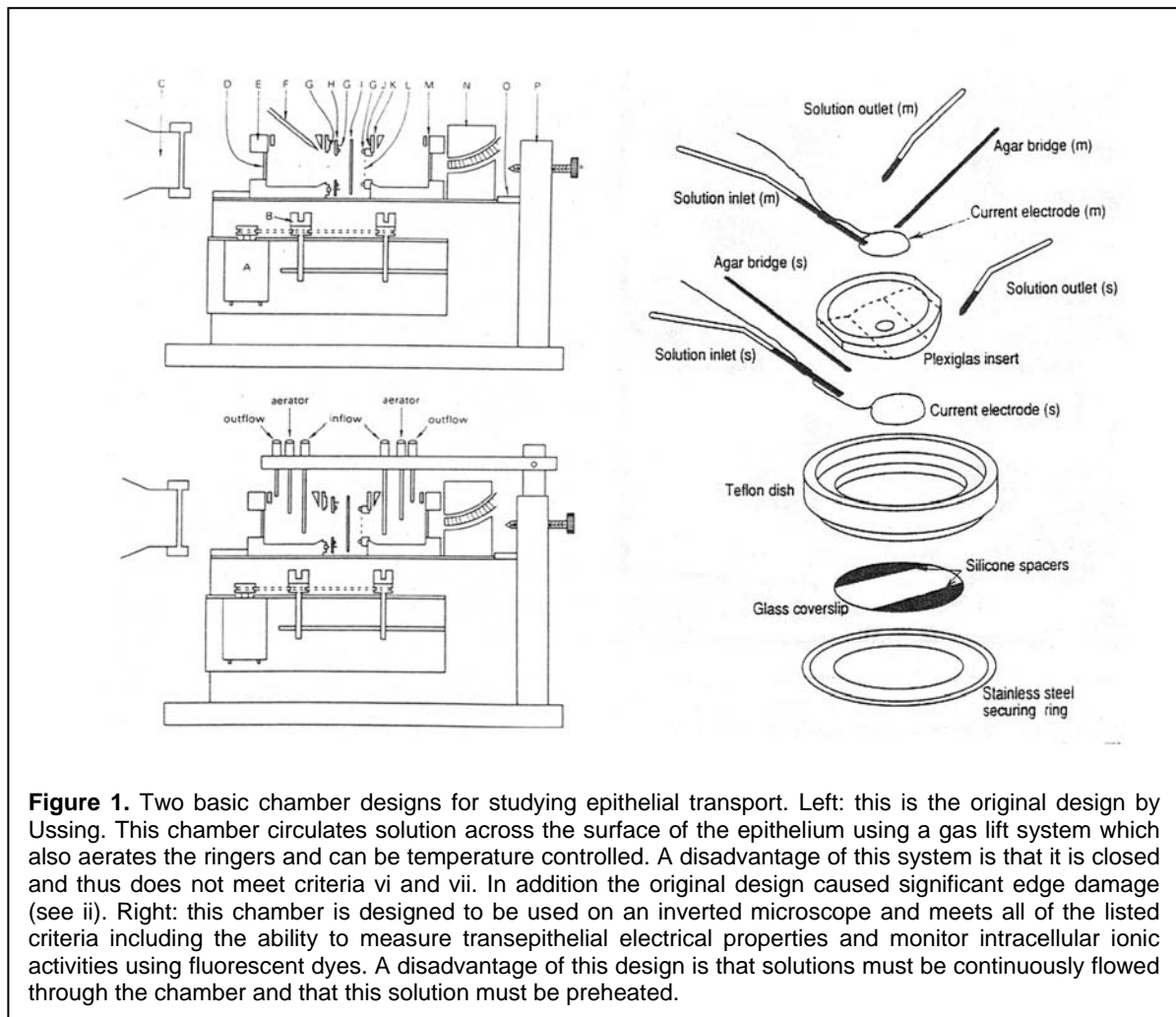
The equipment required for the study of any epithelium (including tissue cultured epithelia) can be divided into three components. The first component is a pair of hemi-chambers (typically termed Ussing chambers) in which the epithelium acts as a partition between the two chambers. Thus the epithelium separates two compartments; the mucosal or luminal compartment and the serosal or blood side compartment. The second component contains the electrodes for sensing the epithelial voltage and for passing current and the third component is comprised of the instrumentation necessary for measuring both voltage and current.

Chambers

Irrespective of the detailed design, useful chambers must have several common features. These include:

- i. **Temperature control:** The chamber must have provisions for controlling the temperature of the mucosal and serosal bathing solutions and for aerating both solutions with the gas mixture of choice.
- ii. **Damage control:** The chamber must have a design which minimizes damage to the cells which contact the inner circumference of the hemi-chamber. If not protected against, such damage (termed edge damage) will result in a measured epithelium resistance which is lower than the native tissue.
- iii. **Support:** The epithelium must be supported on one side by a rigid but permeable structure. This reduces epithelial stretch and the possibility of cell or tight junction damage.
- iv. **Voltage electrodes:** The voltage measuring electrodes must be placed as close as possible to the epithelium. This will reduce the magnitude of the solution series resistance which, if large, can compromise the ability to precisely voltage clamp the epithelium
- v. **Current electrodes:** The current passing electrodes must be placed in the rear of the chambers and as far as possible from the epithelium. This will assure a uniform current density across the epithelium. A non-uniform current density will result in an overestimate of the epithelial resistance.

- vi. Solution changes: Bathing solutions must be easily and rapidly changed without interrupting electrical measurements and without altering the electrical properties of the epithelium.
- vii. Multi-functionality: Ideally, the chamber should be multi-functional. Thus, in addition to measuring transepithelial electrical properties, it should also be constructed to allow for the use of micro- or ion selective electrodes. The former electrodes are essential for determining the individual membrane resistances, while the latter are required to determine membrane ionic permeabilities.
- viii. Additional features might include the ability to measure cell volume and use intracellular fluorescent dyes. Two chamber designs currently in use for the study of epithelial transport are shown below.



Electrodes

Electrodes are an essential component of any electrophysiological set-up since they provide the low resistance interface between the Ringer's solution and the electronic equipment. This section considers the electrodes used to sense the epithelial voltage and to pass a transepithelial current. Although it may seem trivial, careful choice of the electrodes used is important. Some guidelines are listed below.

Choices: One has three choices in voltage measuring electrodes. These are silver/silver chloride (Ag/AgCl) electrodes, calomel electrodes or agar bridges. Which electrode set to use? The answer to this question depends upon the epithelium to be studied, as well as the composition of the solutions bathing the epithelium.

- i. Silver/silver chloride (Ag/AgCl) electrodes can only be used if the epithelium is not sensitive to trace levels of Ag^+ ions (e.g., toad urinary bladder epithelium is very sensitive to trace levels of Ag^+) and if the Cl^- concentration (activity) in the solutions bathing both sides of the epithelium are identical. Unequal Cl^- concentrations will result in an asymmetry voltage difference between the voltage measuring electrodes. This asymmetry voltage will be summed into the transepithelial voltage yielding a voltage artifact (i.e., the measured voltage will be high or low depending upon the ratio of mucosal to serosal Cl^- concentrations).
- ii. If the epithelium is sensitive to Ag^+ or is bathed by solutions containing different Cl^- concentrations, then agar bridges connected to either Ag/AgCl electrodes or calomel electrodes must be used. The advantage of the agar bridge-Ag/AgCl electrode combination compared to calomel electrodes is that they are small and easy to locate close to the epithelial surface. Moreover, they are inexpensive. Agar bridge electrodes must be connected *via* a salt solution to either the Ag/AgCl or calomel electrodes which are connected in turn to the **EC-800**. The same electrode configurations can be used for the current passing limb of the electronics.

Fabrication: Sintered Ag/AgCl pellet electrodes are commercially available from Warner Instruments. Call our offices or see our catalog for our extensive selection. An alternate choice is to use silver wire which has been chlorided by the user. (See Appendix for techniques on chloriding Ag wire).

Agar bridges are easily constructed by heating a mixture of 5% agar with 1M KCl (w/v). While still hot, the agar can be drawn into polyethylene tubing using a syringe or vacuum line. Since the polyethylene tubing is opaque, it is convenient to add a dye (e.g., methylene blue) to the agar/KCl solution. This allows the visualization of any discontinuity in the agar bridge which can cause an overload (lights and audio signal) from the input and/or clamp amplifier.

CAUTION: Current passing electrodes must have a low interface resistance with the solution to work properly.

If the interface resistance is too large, it will limit the current passing capacity of the electronic equipment. Ideally, use of a Ag/AgCl sheet or wire coil in the rear of each hemi-

chamber is sufficient to assure a uniform current density across the epithelium. However, if the tissue is sensitive to trace levels of Ag^+ , a wide bore agar bridge placed at the rear of each hemi-chamber will be required to make a connection between the electrodes and solution. If this technique is necessary, then every effort should be made to keep the length of the bridge short and to fill the bridge with a low resistance solution to maximize the current passing capability of the electronic equipment. A disposable pipette tip partially filled with agar, back filled with a low resistance solution (1M KCl) and coupled to the electronic equipment with a Ag/AgCl wire or sintered pellet works very well.

Electronics

Four important features of an epithelial clamp are described in this section. They are compliance voltage of the current passing amplifier, speed of the current and voltage circuits, series resistance compensation and internal pulse generation.

Compliance voltage of the current passing amplifier (defined as the maximum voltage output of the amplifier) is important if agar leads are used as part of the current passing circuit. Some clamps use amplifiers which have a compliance voltage of 10 volts. If the total resistance of the current passing circuit is 10 k Ω , this means that the maximum current the clamp can pass is +1.0 mA. Although this current level is sufficient for high resistance (tight) epithelia, it most likely will be marginal for low resistance (leaky) epithelia or for epithelia whose resistance has been decreased due to an experimental maneuver. The **EC-800** uses a 110 V current passing amplifier making this an instrument of choice for leaky epithelia. The amplifier also has outputs which can be easily connected to chart recorders, oscilloscopes or computers.

Minimum equipment requirements are:

- i. A current/voltage clamp amplifier: This is essential to record the transepithelial voltage, the transepithelial resistance and the short circuit current (I_{sc}).
- ii. A pulse generator: This device is necessary to either pass a current across the epithelium and record the change in the transepithelial voltage, or to voltage clamp the epithelium and record the current passed to maintain that voltage. Either of these two measuring schemes allows the calculation of the transepithelial resistance.
- iii. A computer with acquisition hardware and software: A well chosen acquisition system can simultaneously display several data channels while digitizing and storing the data in memory for later viewing and analysis. An advantage of using a computer interfaced with the current/voltage clamp is that the data can be immediately analyzed providing feedback on the viability of the epithelium.
- iv. If a computer is unavailable, then a 2-4 channel paper chart recorder and oscilloscope is necessary. The chart recorder will provide a permanent record of the experiment and should have a pen speed fast enough to record changes in transepithelial voltage or current produced by the pulse generator. An oscilloscope is required to display the time dependent changes in transepithelial voltage during a current pulse. In addition, the oscilloscope can be used to determine whether the current/voltage clamp is stable (i.e. it

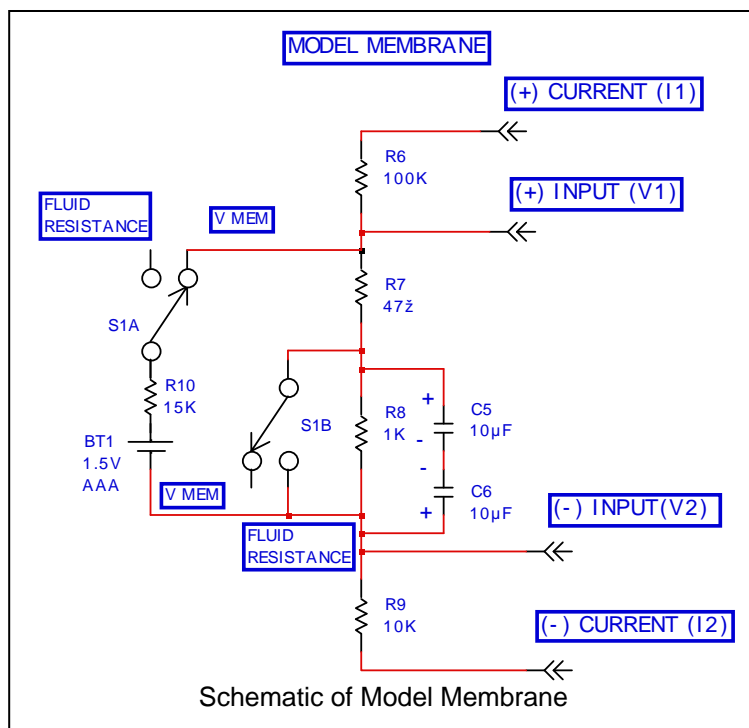
is not oscillating) as well as the response time of the clamp (e.g., is the current or voltage pulse that is being passed square or does it have a finite rise time?). In the **EC-800**, the speed of the clamp (i.e. rate of rise of the current pulse) can be selected on the front panel. A square current pulse is important when estimating epithelial surface area using capacitance measurements (see page 26).

Quick test

The **EC-800** headstage contains a model membrane which can be made functional by selecting the *test* position on the HEADSTAGE SELECTOR switch.

Initial control settings

- Set the HEADSTAGE SELECTOR switch to *off*.
- Set the METER MODE toggle switch to *voltage (mV)*.
- Set the MODE SELECTOR switch to *amplify*.
- Set the polarity switches for INPUT OFFSET, and the COMMAND controls (VOLTAGE and CURRENT), to their respective *off* positions.



After the instrument has warmed up, the digital voltmeters should show a potential close to 0 mV. Completion of the following procedures verifies the proper operation of the voltage clamp.

Monitoring model membrane voltage

- On the HEADSTAGE, set the SELECTOR switch to *test* and the TOGGLE switch to *Vmem*.

The digital panel meter should show a potential in the neighborhood of 100 ± 10 mV, depending on the condition of the battery inside the headstage.

Checking zero voltage clamp

- Change the MODE SELECTOR switch to *V Clamp*.

The panel meter should read 0 V. The membrane is now clamped to zero volts.

Check voltage clamp command

- If + or - is selected on the COMMAND VOLTAGE toggle switch, the 10-turn dial will voltage clamp the membrane at voltages other than zero volts. These clamp voltages should be displayed on the panel METER.
- Return the COMMAND VOLTAGE toggle switch to *off*.

Checking clamp current on panel meter

- Set the METER MODE switch to *current* μA .

The meter now displays the clamp current necessary to clamp the membrane to zero volts.

Fluid Resistance compensation

- Set the MODE SELECTOR switch to *Amplify*.
- Set the METER MODE to *voltage (mV)*.
- Set the HEADSTAGE SELECTOR switch to *test* and the HEADSTAGE TOGGLE switch to *Fluid Resistance*.
- Press the PUSH TO ADJUST button in the FLUID RESISTANCE control block.

The LCD meter should display a *non-zero* value.

- While depressing the PUSH TO ADJUST button, advance the FLUID RESISTANCE POTENTIOMETER until the meter again reads *zero*.

The 10-turn dial on the potentiometer should read approximately 47Ω (the value of the dummy fluid resistor).

- Switch the METER MODE switch to *current*.
- Press the PUSH TO ADJUST button in the FLUID RESISTANCE control block to check the amplitude of the fluid resistance test current, it should be set for $25 \mu\text{A}$.

OPERATION

Procedures are presented for using this experimental set-up to measure the basic electrical properties of epithelia including the transepithelial potential difference, the transepithelial resistance, short circuit current and capacitance.

Connecting the EC-800

This section describes the steps involved in connecting the **EC-800** to an epithelium.

From epithelium to headstage

- Start with the instrument power *off*.
- Set the MODE SELECTOR switch to *Amplify*.
- Set the HEADSTAGE SELECTOR switch *off*.
- Place the voltage measuring and current passing electrodes into the chamber.
- Connected the electrodes to the HEADSTAGE **inputs** using the provided color coded leads. **WARNING: Electrodes connected to V1 (voltage electrode of side 1) and I1 (current electrode of side 1) must be placed in the same bathing solution. Similarly, V2 and I2 must be placed in the same opposing bathing solution.**

From the current/voltage clamp to external monitors

The **EC-800** is a completely self contained unit. However, a permanent record of your data can be exported to a paper chart recorder, oscilloscope or computer via the clamp BNC OUTPUTS (VOLTAGE MONITOR and CURRENT MONITOR in the FLUID RESISTANCE control block).

Checking for asymmetries in the voltage electrodes

Prior to beginning an experiment, asymmetries in the voltage measuring electrodes must be nulled (compensated).

- Set the HEADSTAGE SELECTOR switch to *off*.
- Select *amplify* on the MODE SELECTOR switch.
- Set the METER MODE toggle to *voltage*.
- Place both voltage measuring electrodes (HEADSTAGE **V1** and **V2 inputs**) into the same bath.
- Turn the HEADSTAGE selector switch to *on*.
- If the voltage reading on the meter is non-zero, then use the INPUT OFFSET POTENTIOMETER to adjust the voltage reading to *zero*.

As a rule of thumb, if the offset voltage is greater than 10 mV, then the electrodes must be replaced and the offset readjusted.

- Set the HEADSTAGE SELECTOR switch to *off*.
- Place the HEADSTAGE **voltage measuring electrodes** into their respective chambers.

- Turn the HEADSTAGE SELECTOR switch to *on* and select the desired measuring mode (see below).

Recording modes

The recording mode to be used (voltage clamp or current clamp) depends upon the experimental design. Advantages and disadvantages of different recording modes are listed below.

Voltage clamp: In this mode the transepithelial voltage is set to a specified value and the amplifier passes a transepithelial current to maintain that voltage. The magnitude and polarity of the clamp voltage is set within the COMMANDS control block. Voltage clamp mode can be used, for example, to determine which ions are actively transported by the epithelium (using radio isotopic flux measurements or pharmacological ion transport blockers).

Amplify: In this mode the current passing amplifier is disconnected from the current passing electrodes (HEADSTAGE I1 and I2 inputs). This mode is used to measure any asymmetry in the voltage sensing electrodes. Since the current passing amplifier is disconnected, an external voltage or current applied to the EXT SIGNAL INPUT BNC will not be passed across the epithelium.

Current clamp: In this mode the transepithelial current is typically clamped to *zero* (i.e. there is no net transepithelial current flow), the condition the tissue is exposed to *in vivo*. The steady state current can be varied in magnitude and polarity within the COMMANDS control block. The voltage measured in this mode is produced by the active transport of ions by the epithelium plus the current applied across the epithelium by the current passing amplifier. An advantage of the *zero current* clamp mode that, in the presence of Ag/AgCl wires, the Ag⁺ contamination of the bathing solutions is minimized.

Resistance of the membrane: In this mode, the **EC-800** automatically determines the membrane resistance by passing a bipolar current pulse of 10 μ A (0-2 k Ω range) or 1 μ A (0-200 k Ω range). The resistance is read directly from the panel meter in units of Ω or k Ω (e.g., 103 Ω). If the meter displays all zeros, then select the Ω setting (by pushing the R MEMBRANE RANGE button in the MODE control block). If the meter is blank except for a 1 on the far left, then select the k Ω setting. If the meter's reading does not change, then either the resistance of the tissue is larger than 50 k Ω or there is a high resistance connection between the headstage and the current or voltage electrodes in the chambers.

External timer: This allows the operator to remotely select between current clamp (TTL=*high*) or voltage clamp (TTL=*low*) modes using either a switching box (not provided) or a computer input connected to the EXTERNAL TIMER IN BNC. In this mode, a TTL=*low* signal at the GATE INPUT BNC instructs the EXTERNAL TIMER IN BNC to select between the values set for voltage or current clamp modes. By comparison, a TTL=*high* signal at the GATE INPUT BNC instructs the EXTERNAL TIMER IN BNC to select for *zero current clamp* for current clamp mode or *zero voltage clamp* for voltage clamp mode.

Warning lights

If the voltage between the HEADSTAGE V1 and V2 inputs exceeds 1.2 V there will be an audio warning and visual warning that the input amplifier is saturating. Similarly, a saturation of the clamp amplifier has both audio and visual warnings.

Fluid resistance

The resistance between the voltage measuring electrodes is composed of a series combination of the resistance of the epithelium and the resistance of the fluid bathing the tissue (this latter resistance is sometimes called the series resistance). To determine the epithelial resistance the fluid resistance must be measured and subtracted from the total resistance. Moreover, to voltage clamp a tissue at a specified value, the fluid resistance must be compensated for.

The fluid resistance can be measured and set using one of the following protocols:

Method A

- Connect all electrodes to the chamber and fill with the appropriate Ringers solution in the absence of an epithelium
- Switch the HEADSTAGE SELECTOR switch to *on*.
- Set the METER toggle switch to *voltage*.
- While depressing the PUSH TO ADJUST button in the FLUID RESISTANCE control block, adjust the POTENTIOMETER until the voltage reading on the meter reaches 0 mV.

Method B

- Attach the electrodes and fill chamber as described above.
- Switch the HEADSTAGE SELECTOR switch to *on*.
- Switch the MODE SELECTOR switch to R MEMBRANE mode and read the series resistance from the meter.
- Set the FLUID RESISTANCE POTENTIOMETER to the value read from the meter in the last step.
NOTE: While R MEMBRANE mode disables the FLUID RESISTANCE POTENTIOMETER, the setting will become active in all other operational modes.

Finally, see page 23 for a method of measuring fluid resistance with the epithelium in place (i.e. the fluid resistance adjustment need not be performed before the start of an experiment).

Analyzing the electrical properties of an epithelium

After mounting the epithelium into a chamber, connecting the current/voltage leads to the amplifier, and eliminating asymmetries in the voltage measuring electrode three basic properties of the epithelium can be measured. These are the transepithelial voltage, the transepithelial resistance and the short circuit current (I_{sc} ; a measure of the net active ion transport across the epithelium).

Transepithelial voltage

The first measurement is straightforward and is simply read from the panel meter or from a chart recorder or oscilloscope. One must remember to make sure that there is no asymmetry potential between the voltage measuring electrodes. Since the transepithelial voltage is measured *differentially*, one must know which one of the two voltage measuring electrodes is considered *zero* (or ground).

Transepithelial resistance

The transepithelial resistance can be measured by either passing a current across the epithelium (ΔI) and measuring the resultant voltage change (ΔV under current clamp mode) or by clamping the epithelium to a new voltage (ΔV) and measuring the change in current (ΔI under voltage clamp mode). The resistance is then calculated using Ohm's law which simply states that the resistance is equal to the change in the transepithelial voltage divided by the change in the transepithelial current,

$$R_{meas} = \left(\frac{\Delta V_t}{\Delta I_t} \right) A,$$

where A is the area of the epithelium and the units of resistance are in ohms/cm². Since there is a finite distance between the voltage measuring electrodes and the epithelium, the calculated resistance (R_{meas}) is the sum of the transepithelial resistance (R_t) and the series resistance of the solution (R_s) (i.e. the resistance of the bathing solution between the tissue and each of the voltage measuring electrodes). This series resistance must be subtracted from R_{meas} . To determine the actual transepithelial resistance ($R_t = R_{meas} - R_s$), the **EC-800** can automatically subtract the series resistance (once it has been measured) from R_{meas} .

Series resistance is usually determined by measuring the resistance of the chambers in the absence of an epithelium. Since solutions of different ionic composition have different resistivities, R_s must be uniquely determined for each solution used. Although for high resistance epithelia, series resistance is only a minor correction (e.g. 1-2% of R_{meas}) for low resistance epithelia it can be 50% or greater of R_{meas} . An alternative approach for measuring R_s is offered on page 26.

Short circuit current

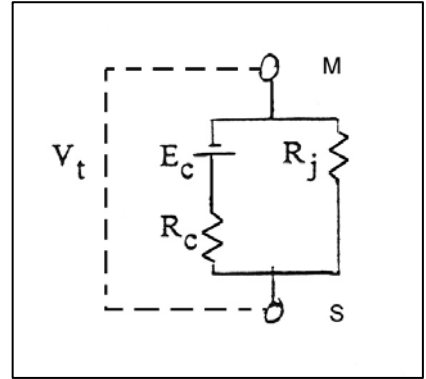
The measurement of the short circuit current (I_{sc}) is deceptively simple. I_{sc} is simply defined as the current that must be passed across the epithelium to reduce the transepithelial voltage to zero. It is the current that short circuits the tissue. I_{sc} is measured by voltage clamping the epithelium to 0 mV and reading the applied current from the panel meter. When performing such measurements, it is essential that one compensates for the series resistance (R_s) by using the FLUID RESISTANCE control block on the **EC-800**.

Equivalent circuit analysis

We now introduced the concept that an epithelium can be modeled as an electrical circuit composed (in the most simple case) of a resistor and a voltage source. The justification for this electrical approach is that many epithelia produce a spontaneous potential even when

bathed on both sides with identical solutions. Therefore, this potential can be represented as a voltage source. Since an epithelium is capable of restricting (resisting) the movement of ions between two compartments, it has resistive properties. Therefore, it can be represented as a resistor. Below is the most simple equivalent circuit of an epithelium based on its morphology.

In the equivalent circuit shown to the right, the cells of the epithelium are represented by a resistor (R_c) in series with a voltage source (E_c), while the parallel tight junctions are represented by a simple resistor. This voltage source (also called the cellular electromotive force, EMF) is a complex function of the conductive properties of the cell membranes and the composition of the ions present in the bathing solution as well as in the cell interior. No voltage source is present in the junction since the tissue is bathed by symmetric solutions. For simplicity, we have left out a series (solution) resistor. Since parallel conductors add, the transepithelial conductance (G_t , the conductance between M, the mucosal solution and S the serosal solution) is given by



$$G_t = G_c + G_j.$$

Since resistance is the inverse of conductance ($G_t=1/R_t$), then the transepithelial resistance is given by

$$R_t = \frac{R_c R_j}{R_c + R_j}$$

and from Ohm's law, the current flow (i) around this circuit is

$$i = \frac{E_c}{R_c + R_j}.$$

Again using Ohm's law we find that the voltage difference across the epithelium ($V_m - V_s = V_t$) is equal to the current flow through the tight junction,

$$V_t = iR_j = \frac{E_c R_j}{R_c + R_j},$$

or through the cellular pathway,

$$V_t = E_c - iR_c = E_c \left(1 - \frac{R_c}{R_c + R_j}\right) = \frac{E_c R_j}{R_c + R_j} = \frac{E_c R_t}{R_c}.$$

Inspection of the above equations reveals three interesting features:

- The magnitude of the transepithelial potential is a function of both the cell and junction resistance, and the magnitude of the cell voltage source.
- The term (E_c/R_c) is equal to the short circuit current and is indeed equal to the current generating capability of the cells.
- At a constant E_c , a decrease in R_c will result in an increase in V_t . Thus as R_c approaches zero, V_t approaches E_c . Similarly, as R_c increases towards infinity, then R_t approaches R_j .

This last relationship can be derived and formalized in the following manner:

$$V_t = \frac{E_c R_t}{R_c}, \text{ rearranging yields } \frac{V_t}{E_c} = \frac{R_t}{R_c}$$

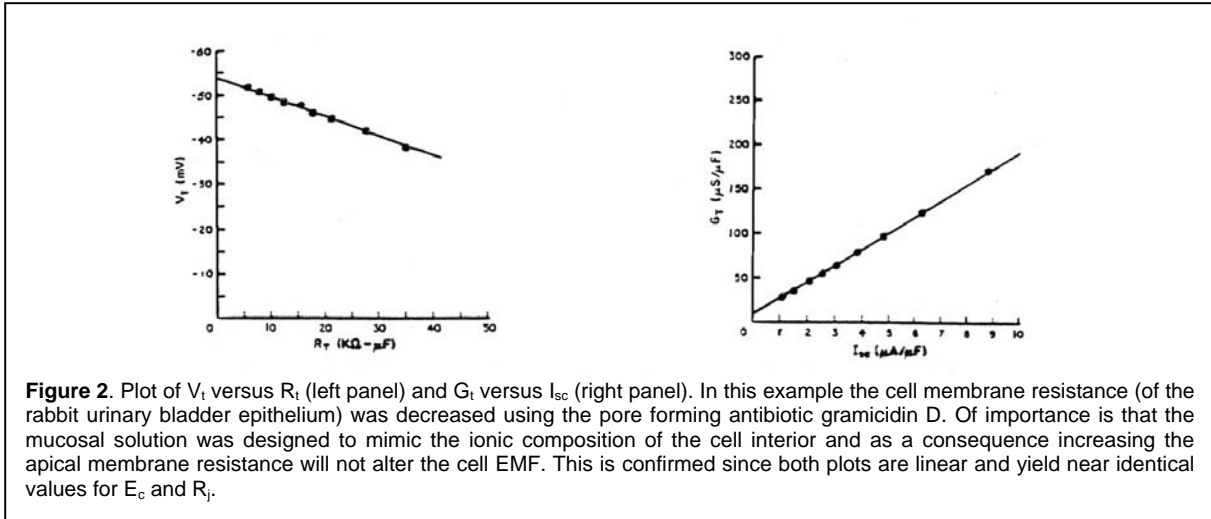
and

$$\frac{1}{R_t} = \frac{1}{R_c} + \frac{1}{R_j}, \text{ rearranging yields } 1 = \frac{R_t}{R_c} + \frac{R_t}{R_j}$$

substituting the above two equations yields

$$\frac{V_t}{E_c} + \frac{R_t}{R_j} = 1.$$

If a perturbation which changes only the cell resistance is performed, a plot of the resulting paired values of V_t and R_t (during this perturbation) yields a linear double intercept in which both R_j and E_c can be determined (see Figure 2, next page). An important question is how does one know that the perturbation has altered only the cell resistance, and not the junction resistance R_j or the cell voltage source E_c ? The best indicator will be that the plot is linear since a curvilinear plot suggests that the assumption of constant R_j and E_c has been violated.



A similar relationship can be derived if the experiments are performed under short circuit conditions. The derivation is shown below.

Recall that

$$G_t = G_c + G_j$$

$$\text{and } I_{sc} = E_c G_c \text{ or } G_c = \frac{I_{sc}}{E_c}.$$

Substituting gives

$$G_t = \frac{I_{sc}}{E_c} + G_j.$$

Using this equation, a plot of the transepithelial conductance (G_t) versus the measured short circuit current (I_{sc}) will have an intercept equal to the junctional conductance (the inverse of the junctional resistance) and a slope equal to the inverse cell voltage (i.e., $1/E_c$). This equation has the same assumption as the previous plotted equation, (i.e. the experimental perturbation only changes the cellular resistance or conductance).

Experience has shown that a plot of V_t vs. R_t is more sensitive to changes in either R_j or E_c than a plot of G_t vs. I_{sc} . The reason for this is that whereas I_{sc} is a measure of only the cellular pathway (it is not affected by the junctional resistance), V_t is a function of both the cellular pathway and the junctional resistance and is thus more sensitive to a change in either parameter. This is illustrated below (Figure 3, next page) in which the experimental conditions were such that both R_c and E_c are changing. Note that although the G_t vs. I_{sc} plot is linear, the plot of V_t vs. R_t is non-linear.

Three approaches have been used to alter R_c in a controlled manner. These are increasing the cell membrane conductance to a given ion using second messenger systems, decreasing the conductance to a given ion using pharmacological blockers, and lastly, artificially increasing the membrane conductance using pore forming agents such as gramicidin D, nystatin or amphotericin B. When using pore forming agents, one must use a mucosa solution which mimics the cell interior (i.e. the potassium content must be high while the calcium, sodium and chloride content must be low). Typically, one performs an equimolar replacement of sodium with potassium and chloride with a large monovalent anion such as gluconate. Such a solution exchange has two advantages. First, since there is low chloride in the bathing solution, cell swelling due to KCl influx is minimized, and secondly, since the

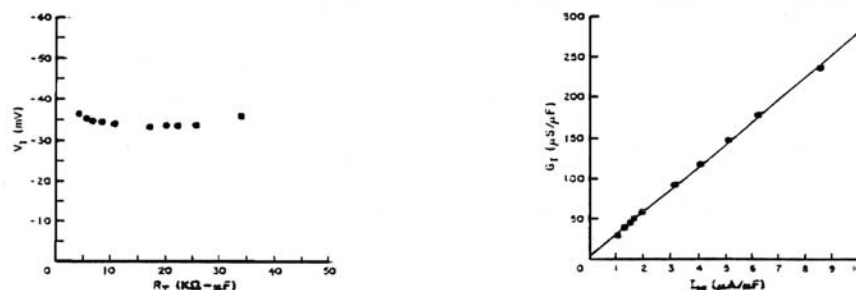


Figure 3. A plot of V_t vs. R_t (left panel) and G_t vs. I_{sc} (right panel). In this example the cell membrane resistance was decreased using gramicidin D. In addition the mucosal solution was selected such that the cell EMF (E_c) changes during gramicidin action on the cell resistance. This is illustrated by the fact that the plot of V_t vs. R_t is non-linear, and indicates that one of the assumptions of the equation has been violated (in this instance E_c is not constant). Note, however, that the plot of G_t versus I_{sc} is reasonably linear even though the assumption of a constant E_c has been violated. To use this method only the resistive (conductive) properties of the cellular pathway must change and the change must be sufficiently large to result in a significant change in the measured parameters, (i.e. V_t , R_t , I_{sc} or G_t). As a consequence, this method is most profitably used on the so called tight epithelia, since a change in the cellular resistance results in a significant change in R_t . In leaky epithelia, where R_t can be 10 fold lower than R_c , large perturbations (in this case a decrease) in R_c must be produced to obtain a reliable (measurable) decrease in R_t .

mucosal and cell ion concentrations are matched, there will be no change in the apical membrane voltage when the pore forming agent is added (i.e. E_c will not be affected as R_c is decreased). Additionally, since the ion concentrations are matched, the value of E_c will be approximately equal to the value of the voltage source of the basolateral membrane.

Determining series resistance and epithelial surface area

In this section we briefly overview how the fluid resistance can be rapidly estimate the with the epithelium in the chamber, as well as how to estimate the surface area of the epithelium. First, we must draw an equivalent electric circuit which describes the epithelium and the fluid resistance in terms of its resistive and capacitative properties. In this circuit, the fluid resistance and the tight junction are modeled as resistors while the apical and basolateral membranes of the epithelium are represented by a parallel arrangement of a resistor and capacitor. The figure below shows this circuit (the tight junctional resistance is not included since it is infinite) and outlines the response of the circuit to a square current pulse. The characteristic equation describing the voltage response to the current input signal is also shown. Note that the voltage response to a square current waveform is time-dependent. Consequently, this response can be analyzed in the time domain.

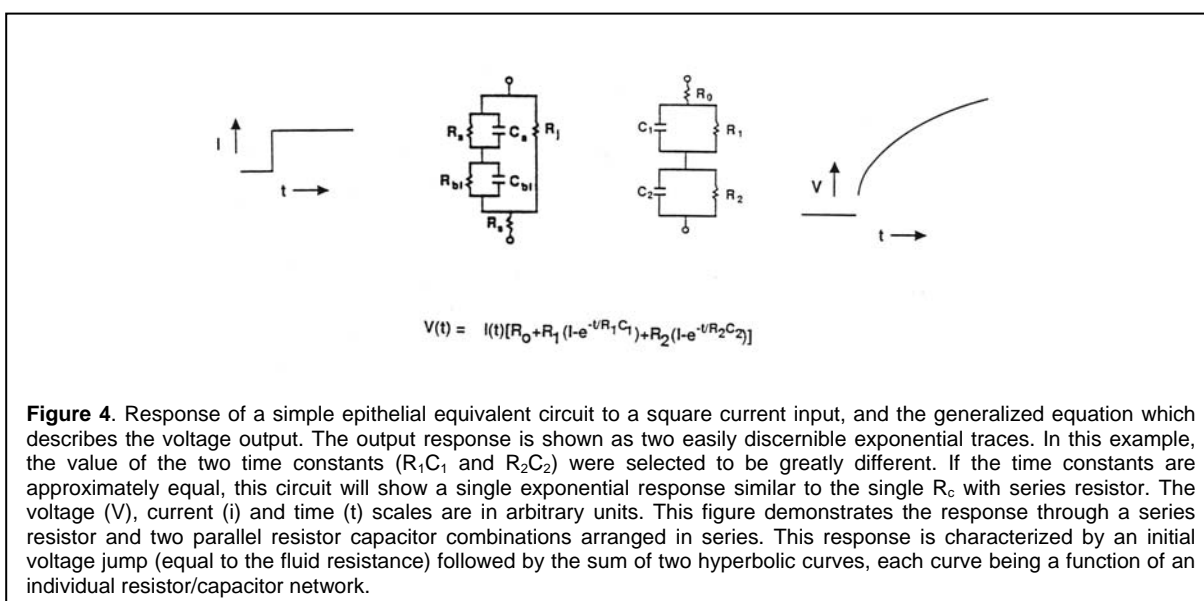


Figure 4. Response of a simple epithelial equivalent circuit to a square current input, and the generalized equation which describes the voltage output. The output response is shown as two easily discernible exponential traces. In this example, the value of the two time constants ($R_1 C_1$ and $R_2 C_2$) were selected to be greatly different. If the time constants are approximately equal, this circuit will show a single exponential response similar to the single R_c with series resistor. The voltage (V), current (i) and time (t) scales are in arbitrary units. This figure demonstrates the response through a series resistor and two parallel resistor capacitor combinations arranged in series. This response is characterized by an initial voltage jump (equal to the fluid resistance) followed by the sum of two hyperbolic curves, each curve being a function of an individual resistor/capacitor network.

MEASURING FLUID RESISTANCE IN THE PRESENCE OF AN EPITHELIAL MEMBRANE

The magnitude of the voltage jump shown in Figure 4 divided by the magnitude of the current step will be equal to the fluid resistance. A square current pulse (the rise time of the current step must be less than 10 μ s) must be used to perform this measurement.

The following protocol can be used to make this measurement:

- Set the MODE SELECTOR switch to *c clamp*.
- Set the CLAMP RESPONSE to *fast*.
- Connect the VOLTAGE MONITOR BNC to a high speed oscilloscope and make sure that the voltage signal is filtered by the oscilloscope amplifier.
- Apply a square current step to the EXT SIGNAL INPUT BNC.

- Measure the magnitude of the initial voltage jump on the oscilloscope. Since the VOLTAGE MONITOR output BNC has x10 gain, divide the measured voltage by 10.
- Divide the measured voltage by the magnitude of the applied current step. The resulting resistance is equal to the fluid resistance.

MEASUREING EPITHELIAL SURFACE AREA

To estimate the epithelial surface area one must analyze the time dependent change in the membrane voltage. The complete protocol for analyzing these voltage transients has been described in detail by Lewis and Demoura (1984). In brief, the "on" voltage response to a square current pulse is digitized (at 100 μ s/point with a resolution of 0.05 mV) and stored in computer memory. This data is next converted to an "off" voltage response by subtracting the time-dependent "on" voltage response from the pre-stimulus voltage. The absolute value of this voltage change ($|\Delta V|$) is stored together with the corresponding time (t; where t=0 is the time at which the current was applied). $|\Delta V|$ is then fit to a sum of exponentials using standard non-linear curve fitting routines.

The number of exponentials used is limited by the equivalent circuit. In general, analysis will yield two voltage terms, which are converted to resistances by dividing the voltage by the amplitude of the current pulse, and two time constants (τ), each of which is the product of a resistor and capacitor. The relationship between the best fit values and the actual epithelial resistors and capacitors depend upon the equivalent circuit model used to represent the epithelium. This is a problem of the non-uniqueness of these equivalent circuits.

FURTHER CONSIDERATIONS

Let us now consider the basic epithelial equivalent circuit model. In Figure 4 we show two circuits. The general form of the equations describing the voltage response of these circuit to a square current pulse are identical. The relationship between the resistors and capacitors in these circuits is described by the following equations (see Lewis and Demoura, 1984):

$$\frac{C_1 C_2}{C_1 + C_2} = \frac{C_a C_{bl}}{C_a + C_{bl}}$$

$$\frac{R_1 + R_2}{R_1 C_1 R_2 C_2} = \frac{R_a + R_{bl}}{R_a C_a R_{bl} C_{bl}}$$

$$\frac{1}{R_1 C_1} + \frac{1}{R_2 C_2} = \frac{R_j + R_a}{R_j R_a C_a} + \frac{R_j + R_{bl}}{R_j R_{bl} C_{bl}}$$

$$\frac{1}{R_1 C_1 R_2 C_2} = \frac{R_j + R_a + R_{bl}}{R_j R_a C_a R_{bl} C_{bl}}$$

Note that a five parameter model (for simplicity we ignore the solution resistance R_s) can be determined by 4 parameters. To curve fit this five parameter model to the impedance data, we must have an independent estimate of one parameter. For example, Clausen *et al.* (1979) measured the ratio of the apical to the basolateral membrane resistances for the rabbit urinary bladder, while Wills and Clausen (1985) independently measured the resistance of the tight junctions using either antibiotics or the sodium channel blocker amiloride.

Of interest is that the first equation demonstrates that the product of the capacitors divided by the sum is independent of the equivalent circuit model. This value is called the effective capacitance and since in most epithelia, C_{bl} is greater than C_a (by about a factor of 5), the effective capacitance is approximately equal to C_a . This relationship has been used to measure the change in membrane surface area as a function of alterations in epithelial transport rate.

APPENDIX

References and recommended reading

- Epithelial Transport: A guide to methods and experimental analysis. Edited by Wills, N.K., Reuss, L., and Lewis, S.A.; Chapman & Hall, London (1996)
- Clausen, C., Lewis, S.A. and Diamond, J.M. 1979. Biophysical Journal. 26: 291-318.
- Crowe, W.E. and Wills, N.K. 1991. Pflugers Archives 419: 349-357
- Lewis, S.A. and Hanrahan, J.W. 1989. Methods in Enzymology 192: 632-650
- Lewis, S.A. and deMoura, J.L.C. 1984. Journal of Membrane Biology 82:123-136.
- Ussing, H.H. and Zerahn, K. 1951. Acta Physiologica Scandinavica 23:111-127
- Wills, N.K. and Clausen, C. 1987. Journal of Membrane Biology. 95:21-35.
- Methods in Enzymology Volume 192. Biomembranes Part W. Cellular and Subcellular Transport Epithelial Cells. 1990. An excellent book that gives detailed methods used to study a variety of epithelia. Includes a discussion on such topics as edge damage and solution resistance.
- Methods in Enzymology Volume 171. Biomembranes Part R. Transport theory: Cells and Model Membranes. 1989. Similar to the above book. Includes good chapters on solution resistance, edge damage artifacts, epithelial impedance, liquid junction potential etc.

Chloriding silver wires

Before using Ag wires as current electrodes, they must be chlorided. New (previously unused) wire should first be cleaned with ETOH before continuing, while previously chlorided wire should have the old chloride coating removed.

Two methods are commonly employed to chloride Ag wire; soaking a clean wire in household bleach or electroplating a clean wire using a voltage source. Both methods are described below.

- A) Soaking in bleach - Simply immerse the wire in full strength common household bleach (Clorox) for 15 to 30 minutes until a purple-gray color is observed. Rinse and use.
- B) Electroplating - Electroplating a silver wire with chloride is achieved by making the wire positive with respect to a solution containing NaCl (0.9%) or KCl (3M) and passing a current through the electrode at a rate of 1 mA/cm² of surface area for 10-15 seconds or until adequately plated (a 1 cm length of 1 mm diameter wire will require approximately 0.3 mA). The color of a well plated wire should be purple-gray. Periodic reversal of the polarity while plating the electrode tends to yield a more stable electrode.

When electroplating a previously plated wire, you may find that it does not plate evenly. Complete removal of the residual silver chloride is usually necessary to effect a uniform coat. Before making the wire positive to the chloriding solution, reverse the polarity for 5 to 10 seconds to remove any remaining chloride that might be left in pits on the wire.. Then proceed as described above.

Adjusting the fluid resistance measurement range

Unless otherwise specified, the FLUID RESISTANCE measurement range is factory set for 0-100 Ω . If required, the FLUID RESISTANCE range can be extended to 1 k Ω by setting jumpers on the main circuit board.

The procedure to make this adjustment is as follows:

- Remove the two Philips-head screws holding the top cover in place and remove the cover.
- Locate the jumper blocks labeled W1-W8. W1-W4 are located on a single block next to the piggy-backed circuit board. W5-W6 and W7-W8 are located on two jumper blocks located towards the front of the main circuit board.
- To set the channel for 1 k Ω operation un-jumper W1, W3, W5, and W7 and jumper W2, W4, W6, and W8.
- To set the channel for 100 Ω operation jumper W1, W3, W5, and W7 and un-jumper W2, W4, W6, and W8.

Connecting the EC-800 to Acquire & Analyze

The basic Acquire & Analyze data acquisition system from Physiologic Instruments is comprised of a DataQ digitizer and the Acquire & Analyze software. Two (2) connecting cables are also supplied; a DB25 (25 pin D connector) printer cable for connecting the DataQ to your computer and a DB37 ribbon cable for connecting the DataQ to an input device.

These components are all that's needed for setup if using an EasyMount multichannel amplifier. If you're using the Warner EC-800, however, then additional components are required: namely a MOD08/A connection box and the associated cables. Generally speaking, you'll make connections from the amplifier to the MOD08/A, and from the MOD08/A to the DataQ.



Hardware connections

1. Begin by installing the Acquire & Analyze software and connecting the DataQ to your computer according to the instructions that came with that equipment.
2. Next, connect the MOD08/A to the DataQ using the supplied DB37 ribbon cable. Use the left side DB37 input on the front of the DataQ.
3. The EC-800 connecting cable has a single DB15 connector on one end and a single DB9 connector on the other end. Connect the DB15 to the associated port on the rear of the EC-800A and the DB9 to a CLAMP input on the MOD08/A.



Software configuration

4. Install and launch Acquire & Analyze
5. Following the instructions on pages 16-17 of the Acquire & Analyze manual, open a new experiment (File/New Experiment) and name it. This will bring up the Experiment Properties dialog box. Make sure the number and location of Active Tissues selected corresponds to the number and location of clamp input on the MOD08/A. (e.g., CLAMP 1)
6. Next, open the Instrument Settings Dialog Box (Setup/Instrument Settings) and verify that the Voltage, Current, and Signal Gains are all set to 10.
7. Click on the ‘Advanced Settings’ button. Set the Instrument Type to EC-800.
8. On the amplifier, set the following switches to the specified settings.

Control	Setting
Offset	<i>off</i>
Command	<i>off</i>
Mode	<i>ext timer</i>
Clamp response	<i>slow</i>
Headstage(s)	<i>Test mode: Vmem</i>

9. In the software, open the Pulse Setup Dialog Box (Setup/Pulse Setup).
10. In the Clamp Mode section, select *Voltage Clamp* and hit *Apply*.
Verify that the amplifier is now in voltage clamp.
Verify that the meter reads voltage ≈ 0 mV and current ≈ 100 -106 uA.
11. In the Clamp Mode section, select *Current Clamp* and hit *Apply*.
Verify that the amplifier is now in current clamp.
Verify that the meter reads voltage ≈ 102 mV and current ≈ 0 uA.
12. In the Clamp Mode section, select *Open Circuit* and hit *Apply*.
Verify that the amplifier remains in current clamp.
Verify that the meter reads voltage ≈ 102 mV and current ≈ 0 uA.
13. In the Clamp Mode section, select *Voltage Clamp*.
Click on *Apply* and exit the Pulse Setup dialog box by hitting *OK*.
14. Open the Reference Setting Dialog Box (Acquire/Reference).
Click on the Reference button and allow the algorithm to fill the table with values.
If you see all white boxes, all yellow boxes, or a combination of white and yellow boxes, then the instrument and software are communicating properly. If you see any red boxes with values greater than 1.0, then contact Warner tech support for assistance.

Specifications

Headstage

Input Impedance:	1 x 10 ¹⁰ Ω shunted by 10 pF
Input Voltage:	±1.5 V maximum
Common Mode Voltage:	±13 V maximum
Common Mode Rejection:	120 dB @ 60 Hz
Leakage Current:	500 pA max.
Offset Voltage Range:	±120 mV

Voltage Clamp

Internal Clamp Potentiometer:	± 100 mV with 10-turn control
External Command:	±1 V
External Command factor:	1 mV/10 mV applied

Current Clamp

Clamp Potentiometer:	±1 mA
External Command:	±10 mA
Command Factor:	1 μA/10 mV applied

Speed: 10 μs measured with model membrane

Compliance

EC-800:	±120 V
EC-800LV:	±15 V

Resistance

Fluid resistance Compensation range:	0-100 Ω standard 0-1 kΩ optional
Membrane Resistance Measurement:	made with a 2 Hz bipolar constant current square wave
Ranges:	0-2 kΩ, injected current = 10 μA 0-200 kΩ, injected current = 1 μA
Panel Meter:	3.5 digit LCD
Voltage Range:	200 mV maximum
Current Range:	2000 μA maximum

Power Requirements

100-130 VAC or 220-240 VAC, 50/60 Hz

Physical Dimensions

EC-800 and EC-800LV: 17.8 cm H x 42 cm W x 25 cm D

Headstage: 7.7 cm H x 7.7 cm W x 5 cm D

Shipping Weight

9.1 kg


Warranty:

Three years, parts and labor

Operating Conditions:

Equipment is intended to be operated in a controlled laboratory environment.	Temperature: 0-40 °C Altitude: sea level to 2000 m Relative humidity: 0-95%
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Certifications

<i>Declaration of Conformity</i>	
<i>CE MARKING (EMC)</i>	
Application of Council Directive: 89/336/EEC	
Standards To Which Conformity Is Declared:	EN55022 Class A EN61000-3-2 EN61000-3-3 EN50082-1:1992 EN61000-4-2 EN61000-4-3 ENV50204 EN610000-4-4 EN610000-4-8 EN610000-4-11
Manufacturer's Name:	Warner Instruments, LLC
Manufacturer's Address:	1125 Dixwell Avenue Hamden, CT 06514 Tel: (203) 776-0664
Equipment Description:	Instrument Amplifier
Equipment Class:	ITE-Class A
Model Numbers:	EC-800, EC-800LV
<i>I the undersigned, hereby declare that the equipment specified above, conforms to the above Directive(s) and Standard(s).</i>	
	Place: Hamden, Connecticut USA
	 Signature:
	Full Name: Burton J. Warner
	Position: President

Declaration of Conformity
CE MARKING (LVD)

Application of Council Directive: 73/23/EEC

Standards To Which Conformity Is Declared:	EN61010-1:1993
Manufacturer's Name:	Warner Instruments, LLC
Manufacturer's Address:	1125 Dixwell Avenue Hamden, CT 06514 Tel: (203) 776-0664
Equipment Description:	Instrument Amplifier Safety requirements for electrical equipment for measurement and laboratory use
Equipment Class:	Class I
Model Numbers:	EC-800, EC-800LV

I the undersigned, hereby declare that the equipment specified above, conforms to the above Directive(s) and Standard(s).

Place: Hamden, Connecticut USA

Signature:



Full Name: Burton J. Warner

Position: President

WEEE/RoHS Compliance Statement

EU Directives WEEE and RoHS

To Our Valued Customers:

Harvard Apparatus is committed to being a good corporate citizen. As part of that commitment, we strive to maintain an environmentally conscious manufacturing operation. The European Union (EU) has enacted two Directives, the first on product recycling (Waste Electrical and Electronic Equipment, WEEE) and the second limiting the use of certain substances (Restriction on the use of Hazardous Substances, RoHS). Over time, these Directives will be implemented in the national laws of each EU Member State.

Once the final national regulations have been put into place, recycling will be offered for those Harvard Apparatus products which are within the scope of the WEEE Directive. Products falling under the scope of the WEEE Directive available for sale after August 13, 2005 will be identified with a "wheelie bin" symbol.

Two Categories of products covered by the WEEE Directive are currently exempt from the RoHS Directive - Category 8, medical devices (with the exception of implanted or infected products) and Category 9, monitoring and control instruments. Most of Harvard Apparatus' products fall into either Category 8 or 9 and are currently exempt from the RoHS Directive. Harvard Apparatus will continue to monitor the application of the RoHS Directive to its products and will comply with any changes as they apply.



- Do Not Dispose Product with Municipal Waste.
- Special Collection/Disposal Required.